



# CRISPR/Cas9 as ONE tool to improve animal models

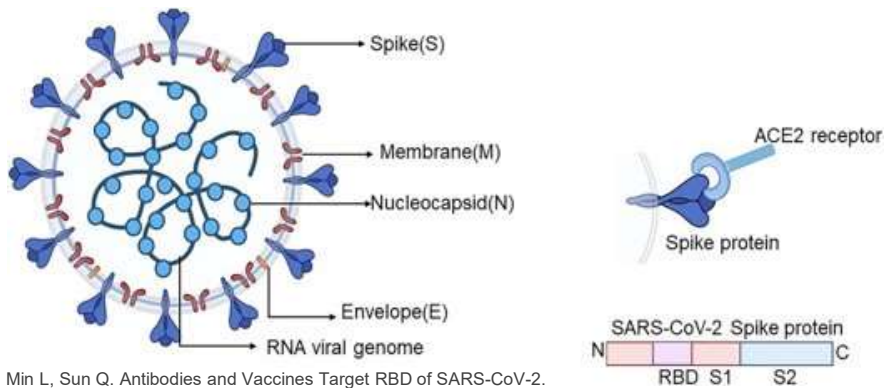
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## Applications for Genetically Engineered Animal Models in Infectious Disease and Vaccine Research

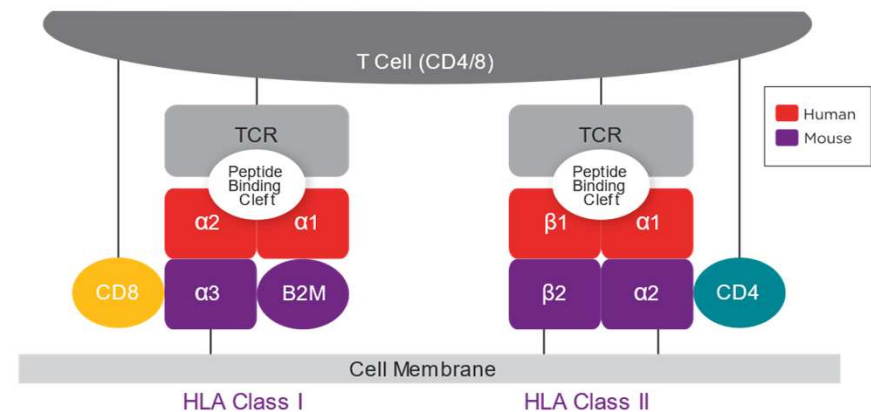
## Conferring Infectivity / Susceptibility



Min L, Sun Q. Antibodies and Vaccines Target RBD of SARS-CoV-2. *Front Mol Biosci*. 2021 Apr 22;8:671633. doi: 10.3389/fmolb.2021.671633. Licensed under CC BY 4.0 (<https://creativecommons.org/licenses/by/4.0/>).

- Humanization of surface proteins necessary for viral entry
- Introduction of human genetic elements that facilitate or enhance infection by pathogens
- Models used for infection studies and measuring efficacy of vaccine therapeutics

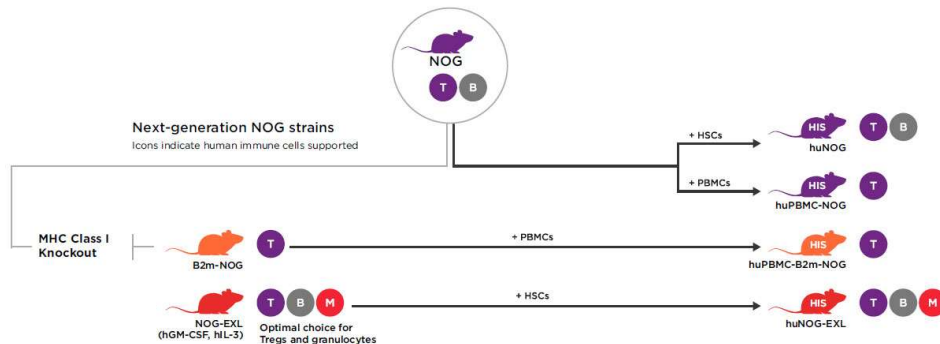
## Humanized Antigen Presentation



- Mice expressing human MHCs (HLAs)
- Transgenes confer human-like presentation of antigens
- Mouse/Human chimeric HLAs facilitate T-cell responses
- Widely used for vaccine development

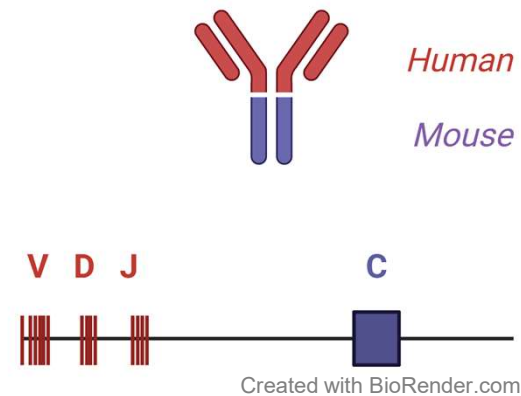
# Applications for Genetically Engineered Animal Models in Infectious Disease and Vaccine Research

## Humanized Immune System Platforms



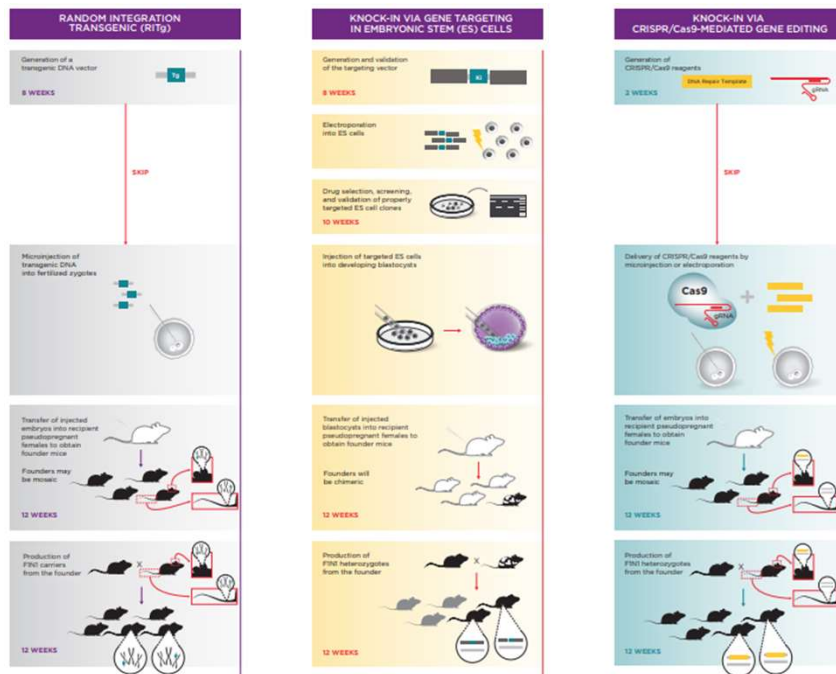
- Genetic inactivation of mouse immune system
- Humanization of cytokines and immune factors supporting development of specific hematopoietic lineages
- Engraftment of human immune progenitor cells
- Examples: NSG mice from Jackson Labs, NOG mice from Taconic

## Humanized Antibody Platforms



- Introduction of engineered transgenes encoding human immunoglobulin repertoires, V(D)J and/or C segments.
- Some platforms retain mouse constant domains and generate human/mouse chimeric antibodies
- Examples: Trianni Mouse, OmniMouse, AlivaMab Mouse

# Genome Engineering Technologies for Model Generation in Rodents



	Random Integration Transgenesis	Gene Targeting in ES Cells	CRISPR/Cas9 Gene Editing
Targeting Method	Zygote Pronuclear Injection	ES Cell Electroporation	Zygote Pronuclear Injection or Electroporation
Type of Integration	Random	Targeted	Targeted
Species	Any <sup>1</sup>	Mouse	Any <sup>1, 2</sup>
Mouse Genetic Backgrounds	Any <sup>1</sup>	Only for strains with established ES cells	Any <sup>1, 2</sup>
Level of Precision	Low	High	Medium, Increasing
Maximum Size of Insertion	≤ 250 kb (BAC)	≤ 180 kb	≤ 5 kb, Increasing
Model Generation Time	≥ 9 months	≥ 12 months	≥ 6 months
Time to First Cohort	≥ 15 months	≥ 18 months	≥ 12 months

<sup>1</sup> any from which zygotes can be harvested and manipulated

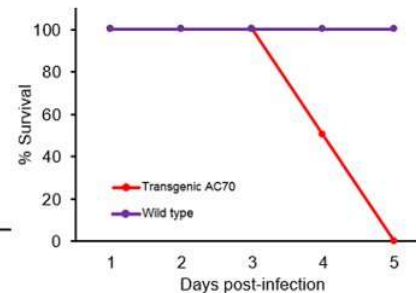
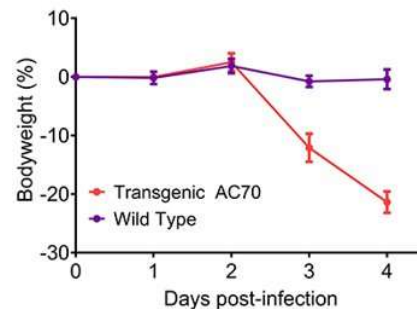
<sup>2</sup> any where genome sequencing data is available

# Example of CRISPR/Cas9 Advantages for Model Development

## Precision, Speed, Accessibility

### Random Integration Transgenic Humanized ACE2 Model

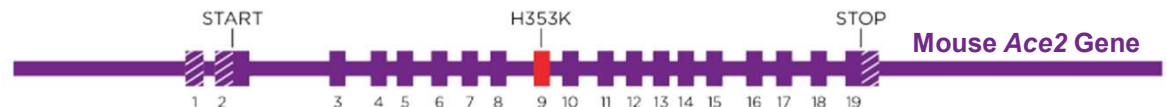
Expressing full length human *ACE2* driven by exogenous promoter



- Infection causes complete lethality
- Unknown location of transgene
- Unknown transgene copy number
- Unpredictable tissue distribution
- Relied on chance of a suitable allele
- Required backcrossing

### CRISPR/Cas9 Point-Mutation Humanized ACE2 Model

Expressing mouse *Ace2* from endogenous locus with humanization of only amino acid H353



- H353K confers SARS-CoV-2 infectivity in mice
- Receptor stoichiometry is maintained
- Receptor retains endogenous signaling
- Receptor is expressed where expected
- No backcrossing necessary
- Precision editing, but 6 months faster than ES cell approach
- Translatable to other strains and species

# Preparedness: Tactics for Rapid & Efficient Model Development

Create generalized work plans for model development suited for specific classes of pathogens

Select the fastest method that provides the scientific requirements with the least risk

Plan a robust and EARLY approach to measuring success and to making a decision on the suitability of the model

Plan ahead the scale that would be needed for immediate studies and how to make accessible to researchers



Genome  
Engineering

Model  
Characterization

Cohort  
Production



Gene editing techniques are rapidly evolving and include emerging classes of programmable nucleases and recombinases with new capabilities

Advances in next generation sequencing platforms are alleviating off-target mutation risk for CRISPR-built models and enabling transgene mapping and characterization

Skip generations in breeding time by using *in vitro* fertilization for rapid colony expansion. Utilize other embryology tools like sperm and embryo cryopreservation to create check points and backup stocks

Apply state-of-the art technologies and industry tricks

# Preparedness: Community-Driven Model Repositories

Reproductive Lifecycle is FIXED. Even with CRISPR/Cas9, the shortest amount of time it takes to generate a *de novo* mouse model and obtain a first experimental cohort is still **~ 1 YEAR**

Anticipate need →

Generate animal models **NOW** and **cryobank** genetic lines

Public Health Organizations  
& Disease Foundations



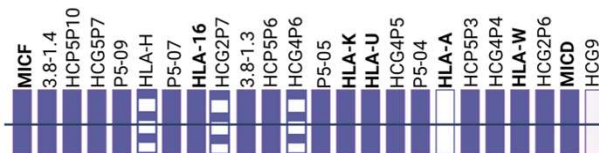
Model Generation Cores &  
Commercial Model Providers



Cryopreserved Models

## Expand Repertoire of HLA-expressing Models

- Create transgenic models for the remaining human HLAs
- Map and characterize HLA transgenes and/or
- Create knock-in versions targeted to safe-harbor loci
- Create engineered transgene(s) encoding full set of HLA



An HLA gene region of Human Chr. 6

Created with BioRender.com

## Create Model Collections for Pathogen Classes

- Humanize common receptors for viral classes
- Introduce susceptibility factors for classes of pathogens or related to human populations
- Create new models on genetic backgrounds permissive to studying infectious diseases

## Clear the path for rapid recovery and distribution

- Create cryorecovery and colony expansion plans
- Anticipate and clear regulatory hurdles to international shipment and global distribution
- Design legal terms to be permissive to sharing with research community



**Thank you for your attention.**

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